
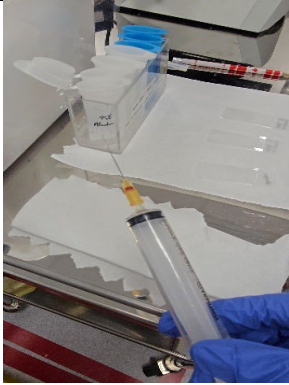
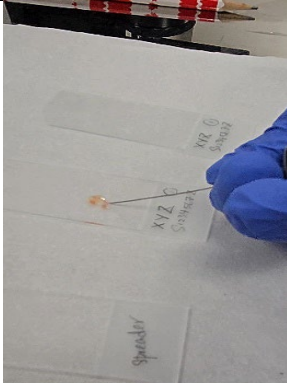
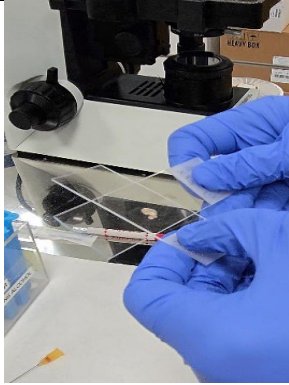
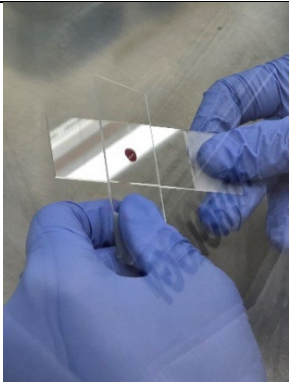
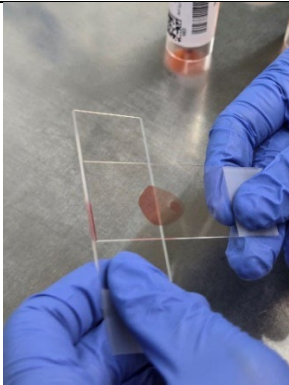

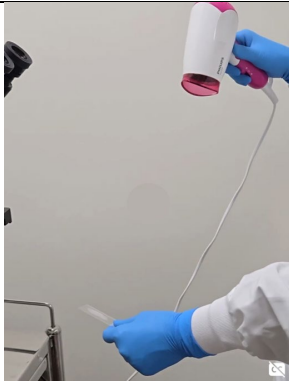
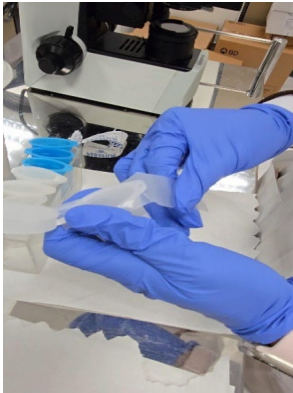
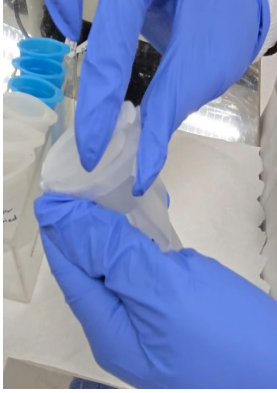
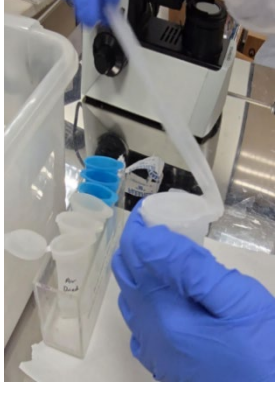



FNA Slide Preparation Workflow Chart

			
<p>1. Label slides with patient initials and IC number.</p>	<p>2. Receive needle from clinician.</p>	<p>3. Detach syringe and fill with air. reattach to needle and expel the specimen onto the slide with the bevel facing down to prevent splash.</p>	<p>4. Hold the slide in your hand. Place a second slide lightly over the specimen to duplicate the material, so you have 2 slides with material.</p>
			
<p>5. Smearing: Use a third slide (spreader) at a 45° angle, gently lower until it contacts the slide with fluid.</p>	<p>6. Smoothly spread the material to form a thin smear. Do steps 5 and 6 for both slides with material. One slide will then be fixed in alcohol immediately (step 7), the other air-dried (step 8).</p>	<p>7. Fix in alcohol: After smearing, immediately place smeared slide in 95% alcohol. Immediate alcohol fixation is necessary.</p>	<p>8. Air-dried smear: Dry using hairdryer on non-heat setting, held 1 foot away</p>

FNA Slide Preparation Workflow Chart

Packing

			
1. Gently stretch parafilm to cover the opening of the slide mailer.	2. Close lid tightly.	3. Pull parafilm to cover the whole lid.	4. Wrap parafilm around 2-3 times to seal.

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Quick Troubleshoot for Staining

PAP with air-drying artifact (Delayed Alcohol fixation)	PAP with good fixation	HC slow air-drying artifact	HC Optimal air-drying
<ul style="list-style-type: none"> Cells swollen Hazy colour 	Immediate fixation <ul style="list-style-type: none"> Cells intact Distinct colour 	<ul style="list-style-type: none"> Pale colour Unclear cell structures 	<ul style="list-style-type: none"> Distinct colour contrast Well defined cell structures
